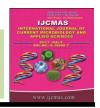


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Toxicity of Some Pesticides to two Important Parasitoids of Lepidopteran Tissue Borers

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ABSTRACT

Keywords

Hymenoptera,
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cephalonica,
Mortality.

Article Info

Accepted: 26 June 2017 Available Online: 10 July 2017 The present study was conducted in the Bio-control laboratory, BCKV located at Mohanpur, Nadia, West Bengal, India to study the toxic effect of some modern insecticides namely cypermethrin 25% EC @ 0.01% a.i., wettable Sulphur 80 WP @ 0.24% a.i., profenofos 50% EC @ 0.05% a.i., in rynaxypyr 20% SC @ 0.006% a.i., spiromesifen 240 SC @ 0.024% a.i. and imidacloprid 17.8% SL @ 0.005% a.i., neemazal 1% EC @ 0.002% a.i. to the pupae of Trichogramma chilonis Ishii (Trichogrammatidae: Hymenoptera) and adults of Bracon brevicornis Wesmael (Braconidae; Hymenoptera). The results showed that Cypermethrin was found to be the most toxic insecticide (65.52% mortality) to the pupae of T. chilonis followed by wettable sulphur (42.7% mortality) and profenofos (37.08% mortality). Toxicity of rynaxypyr, imidacloprid and spiromesifen showed low level of toxicity (27.59%, 25.84% and 26.97% mortality respectively). Neemazal 1% EC was the safest (7.87% mortality). In case of Bracon brevicornis adults, both cypermethrin (0.01% a.i.) and wettable sulphur (0.24% a.i.) were highly toxic (70.35% and 62.84% mortality respectively). Profenofos showed moderate level of toxicity whereas toxicity recorded in imidacloprid, spiromesifen and rynaxypyr was low (32.43%, 32.43% and 35.81% mortality respectively). Neemazal (0.002% a.i) was found to be the safest (20.08% mortality).

Introduction

Pesticides are integral part of present day agriculture. Pesticides are a major factor affecting biological diversity, along with habitat loss and climate change. Wide spectrum insecticides affect a biological process common to many species of insects, so these pesticides are often toxic to nontarget species as well. This is the case with both synthetic and organic insecticides, though organic insecticides usually break down to non-lethal products quickly – a trait that certain synthetic pesticides also have. However, even pesticides that break down quickly are often toxic to non-target species

when exposure is direct. Conversely, if an insecticide is selectively effective only on certain species or families of insects, it is more likely to be safe for beneficial insects and pollinators. Thorough investigation of the effect of insecticides on non-target organisms will help to avoid any potential loss to the environment following their field use. By understanding the difference in toxicity levels of pesticides, a user can minimize the potential hazard by selecting the pesticide with the lowest toxicity that will control the affect pest and not other non-target organisms. Pesticides may affect natural

enemies effectiveness either by causing direct mortality or by influencing their reproduction, behaviour, foraging or movement (Jepson, 1989 and Croft, 1990). Insecticides and acaricides are directly damaging to arthropod natural enemies whereas, the effect of fungicide and herbicides on them may not always be direct. Destruction of natural enemies through indiscriminate use of insecticides has been recognized as one of the major reason for resurgence of many noxious pests/failure of many pest management programmes.

Hymenoptera is the dominant order among all entomophagous insects both numerically and as regard to their successful use in biological control. Among them *Trichogramma* tids (*Trichogramma* tidae; Chalcidoidea), *Trichogramma* are minute parasite wasps that attack the eggs of many lepidopteran insects (Hassan and Abdelgader, 2001). They are the most widely studied and used biological control agents in the world. The genus contains 130 species of which 20 species have been reported from India (Singh and Jahali, 1994).

Bracon is another important parasitoid (Braconidae; Hymenoptera) which are ectoparasitoids of lepidopteran larvae living in concealed places and can easily mass reared in the laboratory (Baker et al., 1995). In the recent past a number of new generation insecticide molecules have been developed which are generally claimed to be rather selective and relatively safer to various groups of non-target organisms. However, a particular insecticide selective to a particular species may be toxic to others. In this backdrop, the present study has been undertaken to study the toxicity of some new generation insecticide molecules to two parasitoids of lepidopteran crop pest namely, Trichogramma chilonis Ishii. and Bracon brevicornis Wesmael.

Materials and Methods

Two important Hymenoptera parasitoids, the egg parasitoid, Trichogramma chilonis Ishii (Trichogramma tidae; Hymenoptera) and the ecto-larval parasitoid, Bracon brevicornis Wesmael (Braconidae; Hymenoptera) were **Bio-control** obtained and reared in Laboratory, Department Agricultural of Entomology, Bidhan Chandra Krishi Viswavidyalaya, West Bengal, India at 27°C temperature and ambient relative humidity.

Insecticides used in the experiment

The experiment was conducted with insecticides viz. Cypermethrin, Rynaxypyr, Spiromesifen, Imidacloprid, Azadirachtin, Profenofos, Wettable Sulphur. The insecticides, their formulations, trade names and address of the manufacturers are presented in table 1.

Laboratory host of the parasitoids

The rice meal moth, *Corcyra cephalonica* Stainton (Galleriidae; Lepidoptera) was used as host for laboratory rearing of both the parasitoids. Sterilised crushed maize grains were used to rear the laboratory host, *C. cephalonica* Stainton.

Mass culture of *Corcyra cephalonica*, laboratory hosts of the parasitoids

Crushed maize grain was sterilized in oven at 100° C for 30 minutes before use to eliminate others insects if any. The sterilized grains, after cooling, were sprayed with 0.2% formalin to prevent the growth of mould as well as to preserve grain humidity, which has lost during heat sterilisation. Then the grains were air dried. The grains were poured into rearing boxes @ 2.5 kg/box and each box was inoculated with 0.5cc *Corcyra* eggs and was closed by placing the lid. The boxes were then

kept in racks. After about 40 days, moths started emerging, that continued for 60 days. Moths were collected daily and were transferred to specially designed oviposition cages placed over a petri plate for egg laying. The eggs were collected every day, cleaned and the scales were removed by blowing by a fan. The collected eggs were either used for further rearing of *Corcyra* or for rearing the egg parasitoid.

Mass culture of *Trichogramma chilonis* Ishii

Eggs of Corcyra, not more than 24h old, were sterilized under UV light (25 watt) for 30 minutes to kill the developing embryo. These were sprinkled uniformly over paper card (15.5cm x 8cm), smeared with a uniform thin layer of 50% aqueous solution of pure gum Arabica with the help of a camel hair brush. Egg cards were then offered to the newly emerged parasitoids for parasitisation inside a glass tube. Super parasitism was avoided by regulating host parasitoid density. The parasitoids were provided with 50% honey solution as food. The temperature of the rearing room was maintained at 27±1°c. The egg card was removed from the tube after 24hrs and a fresh egg card was offered again for parasitisation and the process was continued till 20-25% of the parasioids were alive. Blackening of Corcyra eggs in the card indicated parasitisation and the cards were either kept as such for the emergence of adult parasitoids or utilized or the experiment.

Mass culture of *Bracon brevicornis* Wesmael

Bracon brevicornis was reared on grown up larvae of *C. cephalonica*. About 20 mated females are confined in a glass jar, the mouth of which was secured tightly by muslin sheet and rubber bands. A cotton swab soaked in 50% honey water solution was placed at the bottom of the jar to provide food to the adult

parasitoids. About 30 full grown larvae of Corcyra are placed between two layers of facial tissue paper and were placed over the muslin sheet covering the mouth of the jar. The tissue papers are again covered with a muslin sheet and fastened tightly with rubber bands. After 24 hrs, the tissue papers containing parasitized larvae were removed and kept in plastic containers. The body remnants of Corcyra larvae were removed after pupation of *B. brevicornis* and the pupae were then kept in glass jars measuring 12.5cm x 10cm (dia) for the emergence of adult parasitoids.

Preparation of different concentrations of the insecticides

The toxicity of selected insecticides, each at two concentrations, was evaluated on the pupal and adult stages of the parasitoids. The desired concentrations were prepared by mixing measured quantities of insecticides with fresh tap water. The treatment details are provided in table 2.

Toxicity of different insecticides on the pupae of *Trichogramma* spp.

For the experiment, parasitized egg cards were kept for development of the parasitoids inside the host eggs. On the fifth day after parasitisation when all the parasitized eggs have turned black indicating that, the parasitoids inside have entered pupation, the egg cards were made into pieces, each piece containing at least 50 parasitized eggs. These small cards were dipped in insecticidal solutions, removed immediately and dried under fan. In control treatment, egg cards were dipped in water. The egg cards were then kept separately in glass vials to allow the emergence of adult parasitoids from the insecticide treated host eggs. The mouth of the vials was covered with muslin cloth tied with rubber bands. The pupal mortality was recorded after adult emergence and the pupae

from which no adult has emerged were considered as dead. Each treatment was replicated three times and observation was taken under a binocular Stereo Microscope on 30 parasitized eggs in each replication (10 from three different locations of the card)

Toxicity of insecticides on the adults of *B. brevicornis* Wesmael

Insecticidal solutions were prepared as mentioned earlier. Field collected castor leaves were cut into pieces measuring 12.5cm x 2.5cm (dia), dipped in insecticidal solution and were dried under fan. The leaves were then placed inside glass tubes measuring 12.5cm x 2.5cm (dia) covering the entire inner surface of the tube and ten numbers of newly emerged *B. brevicornis* adult females were released within the tube. The mouth of the glass tubes was plugged with fine cloth and tied with rubber bands. Mortality was recorded after 24 hrs and the moribund adults were considered as dead. Each treatment was replicated thrice.

Statistical methods

From the observed % mortality in different insecticidal treatments and untreated control, corrected % mortality of the pupae was calculated by using Abbott's formula (1925).

 $P' = P - C / 100 - C \times 100$

Where,

P'= Corrected % mortality P=Observed % mortality C= % mortality in control

Results and Discussion

The toxicity of seven pesticides (insecticides and acaricides) to the pupae to *Trichogramma Chilonis* Ishii (Braconidae; Hymenoptera) and adults of *Bracon brevicornis* Wesmael (Braconidae; Hymenoptera) evaluated in the laboratory showed the following results

Toxcity of pesticides to *Trichogramma* chilonis Ishii. Pupae

Data presented in table 3 showed that among the pesticidal treatments cypermethrin 25% EC at the rate of 0.01% a.i. was most toxic causing 65.52% mortality of the treated pupae and this treatment was statistically more toxic than all other pesticides used. Similarly, Samanta et al., (2009) found cypermethrin to be highly toxic to Trichogramma chilonis which corroborates the fineness of the present investigation. Wettable Sulphur 80 WP at the rate of 0.24% a.i. caused 42.7% mortality of the pupae which was statistically at par with profenofos 50% EC at the rate of 0.05% a.i. (37.08% mortality). These two treatments, however, showed significantly higher toxicity than treatment with rynaxypyr, spiromesifen, imidacloprid and neemazal. Mandal and Panja (2006) reported that profenofos 0.0625% caused total mortality of Trichogramma chilonis. During the present investigation, profenofos however, 0.05% a.i. moderately toxic to the parasitoids. Pupal mortality recorded in rynaxypyr 20% SC at the rate of 0.006% a.i., spiromesifen 240 SC at the rate of 0.024% a.i. and imidacloprid 17.8% SL at the rate of 0.005% a.i. was 27.59%, 25.84% and 26.97% respectively which were statistically at per among them. Karam 2007) found rynaxypyr 20% SC to be the safest to the pupae of T. chilonis. Jayashree et al., (2002) found imidacloprid to be safest to Trichogramma chilonis whereas Nasreen et al., (2004) reported imidacloprid as toxic to the parasitoids. On the other hand, Basappa (2007) reported imidacloprid as moderately toxic. During the present investigation, however, imidacloprid moderately found be to Trichogramma chilonis. Differences in the experimental methodology conditions. adopted, age of the parasitoids exposed and strains of the parasitoids may also be responsible for these observed differences by the authors.

Table.1 List of the insecticides along with their formulation and sources

SL. No.	Insecticides	TradeNames	Formulation	Manufacturer With Address
1.	Cypermethrin	Super Killer	25% EC	DhanukaAgritech Ltd
2.	Rynaxypyr	Coragen	18.9 % w/w SC	M/S E.I. DuPont India Pvt. Ltd.
3.	Spiromesifen	Oberon	240 SC	Bayer crop science Limited
4.	Imidacloprid	Confidor	17.8 % SL	Bayer crop science Limited
5.	Azadirachtin	Neemazal	1 % EC	E.I.D parry (India) ltd.
6.	Profenofos	Carina	50% EC	PI Industries Ltd.
7.	WettableSulphur	Sulfex	80 WP	Excel industries pvt.ltd.

Table.2 Name of chemical with their concentrations

S. No	Insecticides	Doses(ml/litre)
1	Cypermethrin (25% EC)	0.4
2	Rynaxypyr (18.9 %w/w SC)	0.3
3	Spiromesifen (240 SC)	1
4	Imidacloprid (17.8 SL)	0.3
5	Azadirachtin (1 % EC))	2
6	Profenofos (50% EC)	1
7	WettableSulphur (80 WP)	3(gm)

Table.3 Percent mortality of Trichogramma chilonis Ishii

S. No	Insecticides	% Pupal mortality
1.	Cypermethrin 25% EC @ 0.01% a.i.	60.22 (54.04)
2.	Rynaxypyr 20% SC @ 0.006% a.i.	27.59 (31.64)
3.	Spiromesifen 240 SC @ 0.024% a.i.	25.84 (30.47)
4.	Imidacloprid 17.8 SL @ 0.005% a.i.	26.97 (31.24)
5.	WettableSulphur 80 WP @ 0.24% a.i.	42.70 (40.77)
6.	Profenofos 50% EC @ 0.05% a.i.	37.08 (37.45)
7.	Neemazal 1% EC @ 0.002% a.i.	7.87 (16.21)
F test	Significant	
SE(m)±	1.62	
CD at 5%	4.90	

Figures in the parentheses are angular transformed values

Table.4 Percent mortality of Bracon brevicornis Wesmael

S. No	Insecticides	% Adult mortality	
1.	Cypermethrin 25% EC @ 0.01% a.i.	76.35 (60.98)	
2.	Rynaxypyr 20% SC @ 0.006% a.i.	35.81 (36.60)	
3.	Spiromesifen 240 SC @ 0.024% a.i.	32.43 (34.65)	
4.	Imidacloprid 17.8 SL @ 0.005% a.i.	32.43 (34.65)	
5.	WettableSulphur 80 WP @ 0.24% a.i.	62.84 (52.46)	
6.	Profenofos 50% EC @ 0.05% a.i.	44.59 (40.69)	
7.	Neemazal 1% EC @ 0.002% a.i.	20.08 (20.08)	
F test	Significant	<u> </u>	
SE(m)±	2.75		
CD at 5%	8.34		

Figures in the parentheses are angular transformed values

Neemazal1% EC at the rate of 0.002% a.i. resulted in 7.87% mortality only and was proved to be the safest to the parasitoids.

Toxicity of pesticides to *Bracon brevicornis* Wesmael adults

The corrected mortality of Bracon brevicornis wesmael adults after 24 hrs exposure to the pesticides are presented in table 4. Here also cypermethrin 25% EC at the rate of 0.01% a.i. was the most toxic insecticides (70.35% mortality), followed by wettable sulphur 80 WP at the rate of 0.24% a.i. (62.84% mortality). Reports relating to the toxicity of the pesticides to Bracon brevicornis are verylow. However, Guddewar (1992) found cypermethrin to be toxic to brevicornis whereas Chaturvedi (2005)reported cypermethrin as slightly harmful to it. Profenofos 50% EC at the rate of 0.05% a.i. ranks third in respect of toxicity to the parasitoids. This treatment, however, fell to show insignificant difference from the rynaxypyr 20% SC at the rate of 0.006% a.i. (35.81% mortality). Mandal and Panja (2006) observed total mortality Bracon of brevicornis adults even at 0.037% a.i. of profenofos. During the present investigation, however, profenofos at the rate of 0.05% a.i. was relatively safe to the natural enemies. Both spiromesifen 240 SC at the rate of 0.024% a.i. and imidacloprid 17.8% SL at the rate of 0.005% a.i. produce 32.43% mortality of adults and these two treatments were statistically similar to rynaxypyr 20% SC. neemazal 1% EC at the rate of 0.002% a.i. was the safest causing 20.08% mortality only to the treated adults. It is evident from the results pesticides. that among treated cypermethrin 25% EC at the rate of 0.01% a.i. was highly toxic to both the natural enemies. Wettable sulphur 80 WP at the rate of 0.24% though showed high level of toxicity to the adults' of B. brevicornis show moderate level of toxicity in the pupae of T.chilonis.

Wettable sulphur, an acaricide, results in moderate to high level of mortality to both the natural enemies, however, the result could not be compared with any earlier experimental data. due lack to of information.Profenofos50% EC at the rate of 0.05% a.i., rynaxypyr 20% SC at the rate of 0.006% a.i., spiromesifen 240 SC at the rate of 0.024% a.i. and imidacloprid 17.8% SL at the rate of 0.005% a.i showed moderate level of toxicity to both the parasitoids. Neemazal1% EC at the rate of 0.002% a.i. showed exceptional safety to the natural enemies. It is also evident from the result that toxicity of the pesticides to B. brevicornis adults was slightly higher than the pupae of the T. chilonis. Though pharate adults are generally believed to be highly sensitive to the pesticides the host egg shell may act as a protective barrier thereby resulting in low mortality of the pupae. Moreover, the treatment was given just after pupation of the parasitoids inside host eggs and the young pupae may not be as susceptible as the older ones or the pharate adults.

The results from this experiment helps to conclude that neemazal (0.002% a.i) can safely be used in the fields where both the parasitoids are active whereas cypermethrin should be applied with much caution. Rynaxypyr, spiromesifen, imidacloprid and also profenofos can be utilized without much harm to the natural enemies. Wettable sulphur should be applied with due care particularly in situations where *Bracon brevicornis* is active.

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